MINI REVIEW

Open Access

Old drugs, new challenges: reassigning drugs for cancer therapies



Paulina Czechowicz¹, Anna Więch-Walów¹, Jakub Sławski¹, James F. Collawn² and Rafal Bartoszewski^{1*}

*Correspondence: rafal.bartoszewski@uwr.edu.pl

¹ Department of Biophysics, Faculty of Biotechnology, University of Wroclaw, F. Joliot-Curie 14a Street, 50-383 Wroclaw, Poland ² Department of Cell, Developmental and Integrative Biology, University of Alabama at Birmingham, Birmingham, USA

Abstract

The "War on Cancer" began with the National Cancer Act of 1971 and despite more than 50 years of effort and numerous successes, there still remains much more work to be done. The major challenge remains the complexity and intrinsic polygenicity of neoplastic diseases. Furthermore, the safety of the antitumor therapies still remains a concern given their often off-target effects. Although the amount of money invested in research and development required to introduce a novel FDA-approved drug has continuously increased, the likelihood for a new cancer drug's approval remains limited. One interesting alternative approach, however, is the idea of repurposing of old drugs, which is both faster and less costly than developing new drugs. Repurposed drugs have the potential to address the shortage of new drugs with the added benefit that the safety concerns are already established. That being said, their interactions with other new drugs in combination therapies, however, should be tested. In this review, we discuss the history of repurposed drugs, some successes and failures, as well as the multiple challenges and obstacles that need to be addressed in order to enhance repurposed drugs' potential for new cancer therapies.

Keywords: Drug discovery, Drug repurposing, Cancer, Off-label use, Pharmaceutical development

Introduction

Despite decades of ongoing efforts in the war against cancer, current therapeutic options often remain insufficient [1, 2]. The unresolved challenge is the complexity and intrinsic polygenicity of neoplastic diseases, which cannot be addressed by targeting a single molecular target. A combination of different treatment modalities is usually required to achieve optimal outcomes that often depend on the individual patient and cancer type. The development of effective and sophisticated treatment strategies is currently fueled by rapid progress in basic biomedical research as well as the evolution of surgical, radio-therapeutic, and immunological approaches.

Nevertheless, the safe application of antitumor drugs is limited by the off-target effects of most current drug strategies. Additionally, their prolonged use can result in modified tumor occurrences and resistance, as well as their ability to promote cancer stem cells [3]. Furthermore, with the acknowledgment of inter- and intra-tumoral heterogeneity



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http:// creativecommons.org/licenses/by/4.0/.

and the dynamic and complex interactions within the tumor microenvironment (TME) [4–7], the necessity of simultaneously targeting a variety of different and complex signaling pathways has becomes evident. Consequently, complex combination or multimodal therapies are required [8]. Such approaches could enhance outcomes through their synergistic mechanisms of action by targeting the different properties of cancer cells or the TME. This creates a demand for alternative approaches in drug development [9, 10].

The amount of money invested in research and development (R&D) required to introduce a novel FDA-approved drug is continuously and dramatically increasing, while the likelihood for cancer drug approval in phase I clinical trials remains the lowest of any drug type at 6.7% [10–12]. This limitation in R&D productivity is termed as 'Eroom's Law' [11]. Scannell et al. proposed four problems: (1) 'the Beatles' problem'. In other words, developing new drugs that were significantly more efficient, safer, and improved over existing (and usually cheaper) therapies was like finding a music group better than the Beatles [13]; (2) the 'cautious regulator' problem is the increasing safety and formal requirements by the regulatory agencies [14–17]; (3) the 'throw money at it' problem occurs when the investment strategies try to overrate a potential new drug in order to improve the company competitiveness [18]; (4) and finally the 'basic research–brute force' bias that improvement of basic research and screening technology will always translate into effective drug discovery [18].

These limitations cannot be easily solved or controlled. Other challenges, however, are being addressed through the development of in vitro pharmacological profiling of drug candidates [19] and the integration of increasing basic research information into drug development pipeline design. This is particularly true in the human genome-wide association studies (GWASs) [20]. Furthermore, the careful consideration of the risk–benefit balance by regulatory agencies is also clearly important [21].

One strategy drug development limitations is the repurposing of already existing, clinically approved drugs for combinatory anticancer treatments [22-24]. This approach allows for the expedited development of novel therapies at a fraction of the costs and risks associated with novel drug discovery given that the safety profiles of these molecules have already been established. Furthermore, drug repurposing strategies are supported by progress in understanding both the mechanisms of human pathologies and the long-term consequences of these "old" drugs' applications and by the continuous advancements in targeted drug delivery. Notably, the majority of currently marketed drugs have the ability to interact with more than one target, and occasionally include those that could benefit cancer patients [25-27].

Indeed, opportunistic findings led to the very first chemotherapeutics, chlorambucil (Leukeran) and busulfan (Myleran), which are still used to treat chronic lymphocytic and myeloid leukemias (CLL and CML) [28–30]. These drugs originated from 'mustard gas' and could be regarded as repurposed chemical weapons [31–33]. Nowadays, repurposing strategies often aim to substitute cytotoxic therapeutics with cytostatic ones, such as metformin and thalidomide, originally used to treat diabetes and inflammation, respectively [33]. Drug repurposing is not only limited to finding new uses for drugs initially developed for other diseases (including generics, on-patent drugs, and failed molecules), but also includes the original cancer drugs reassigned for different types of cancer or in different combinations.

Repurposing drugs can also be faster and less costly. For example, despite the need to test the new indications of repurposed drugs in later-phase clinical trials and obtaining approvals, knowing their safety profiles and pharmacokinetics/pharmacodynamics reduces the risk of failure [34, 35]. Although, the vast majority of molecules that undergo clinical trials do not make it to the clinic [36], some of those that fail at late stages could still be good candidates for repurposing and turn financial losses into potential successes.

In summary, drug repurposing strategies could improve the outcomes of modern mono and combinatory antitumor therapies. However, transitioning repurposed drugs into clinical application is accompanied by numerous commercial, technological, and regulatory obstacles. This short narrative review summarizes and discusses both the advantages and challenges associated with the repurposed drug component of antitumor therapeutic strategies.

Strategies for drug repurposing

Drug repurposing and is also known as drug repositioning or drug reprofiling. This classification can be further expanded to include failed drugs—compounds that have entered clinical trials but did not succeed due to unsatisfactory efficacy against the initial indication [37, 38]. All these compounds can be further divided into patent-protected (both approved and failed) and off-patent (generic) drugs. Obviously, the majority of research information comes from the latter group [37, 39, 40].

Although many successful drug reassignments have had rather serendipitous backgrounds, these favorable outcomes drive the optimistic and further development of systematic, dedicated strategies [41]. The simplest classification of drug discovery, including repurposing aspects, is divided into two basic lines of action: target-based drug discovery and phenotypic drug discovery [29]. The first approach starts by defining the molecular target underlying the pathology and aims to design dedicated drugs. For example, tamoxifen, which was initially aimed to be a contraceptive, was repurposed for the treatment of breast cancer since it was found to efficiently inhibit the estrogen receptor [42]. In contrast, phenotypic drug discovery ignores a specific drug target or hypothesis about its role in disease and instead tests candidates for desirable biological activity in 'physiologically relevant' systems. For instance, during a phenotypic screen for cell proliferation, it was found that auranofin, originally an anti-arthritic medication, effectively and selectively targets gastrointestinal stromal tumors, including imatinib-resistant ones [43, 44].

Drug repurposing strategies can also be classified into three main groups: targetcentric, drug-centric, and disease-centric approaches [23]. Each of these strategies has its success stories, limitations, and advantages, and consequently, a dedicated group of supporters. The drug-centric approach focuses on identifying new indications for existing drugs, which can include expanding the existing license or patent towards novel off-label use of the compound in new medical conditions or groups of patients. This strategy is often applied to investigational or failed drugs that faced Eroom's Lawrelated limitations in their initial assignment pipelines [23]. For example, valproic acid, originally indicated for bipolar disorder, has an off-target interaction with histone deacetylase 2, a protein that plays a role in many types of cancers. This has led to testing the repositioning of this drug for the treatment of neoplastic conditions such as familial adenomatous polyposis [45].

Drug-centric repositioning can be considered the least direct approach because the drug is only linked to a novel indication via the discovery of a target that is already established for this indication. Thus, a precise characterization of drug-target interactions is required to propose a novel repositioning hypothesis. The most common technical approaches for drug-centric repositioning are structure-based computational methods like molecular docking [46], pharmacophore modeling algorithms [47], protein–ligand interaction profile similarity testing [48], and machine learning approaches [49, 50]. However, since the drug needs to be repurposed to a novel target or disease, a structural model describing the binding mode of the drug to its original targets needs to be well defined. Furthermore, machine learning approaches are limited by the completeness of information in the databases that are utilized [20, 23, 51].

The target-centric approach matches a new indication without a treatment with an established drug and its known target. The old and new indications typically differ quite significantly. Complementary to a disease-centric approach, target-centric repositioning builds on a novel link between a new indication and an established target. It involves investigating the specific molecular targets implicated in the pathology of a disease and uses an existing drug proven to modulate those targets. For example, azacitidine, a potent inhibitor of DNA methyltransferases, was originally dedicated to treating myelodysplastic syndrome [52] and was later adapted for treating patients with acute myeloid leukemia and chronic myelomonocytic leukemia [52]. This approach is particularly useful when seeking to repurpose drugs to treat rare diseases.

Finally, the most effective approach so far is the disease-centric approach, which involves re-profiling drugs among different types of a disease, such as two types of cancer [23]. It involves identifying diseases with homologous underlying biological mechanisms and similar guiding principles to the indicated original drug treatments. For example, a drug developed to treat psoriasis could also treat other diseases with uncontrolled cell growth, such as cancer. In the case of cancer, these guiding principles are summarized in the Hallmarks of Cancer [53, 54]. Since these key hallmarks of malignancy are not regulated by a single signaling pathway [55], the pathways responsible for a cancer phenotype underlie the pathomechanisms of many other non-oncological human diseases. This opens the possibility of repurposing drugs towards novel anticancer therapeutics and supports agents in combinational approaches [56]. Notably, the diversity and complexity of the hallmarks of oncogenesis provide a strong rationale for using multiple drugs to obtain satisfactory therapy outcomes [40], while drug repositioning could significantly reduce costs and increase the availability of such novel therapies [56, 57].

Cancer complexity as a target of repurposed drugs

With the progress in understanding the molecular mechanisms related to oncogenesis, cancer progression, and treatments, more distinct attributes and signaling pathways are now recognized as crucial for various neoplastic diseases. Indeed, the initially proposed eight Hallmarks of Cancer have now been extended to fourteen [53, 54]. Although this diversity and complexity of neoplastic disease remain a therapeutic challenge, it also provides a strong rationale for drug repurposing. It is important to note that these

hallmarks often result from the crosstalk between different signaling pathways, some of which are deregulated in cancer cells, while others compensate for this [58–62]. For example, cancer cells often efficiently avoid unfolded protein responses, hypoxia, and/or oxidative stress-related cell death signals and favor the proadaptive ones [63–67].

Nevertheless, the complexity of such signaling often limits the ability to selectively target cancer-specific signals without interfering those that are therapeutically desired. For example, inositol-requiring enzyme 1 (IRE1) activity, which is of great interest as a target for glioblastoma and triple-negative breast cancer, can lead to the accumulation of proadaptive signals. Whereas, at the same time, this enzyme can also support cell death signaling [7, 68–70]. Thus, despite the availability of both IRE1 inhibitors and potentiators, their translation into the clinic remains a challenge. Furthermore, healthy cells are often exposed to stress-inducing factors, with chemotherapeutics being one of them [71], and thus such stress-oriented therapies also carry the risks of adverse effects.

Chronic proliferation, a fundamental feature of cancer cells, often results from a network of deregulated signaling pathways and growth factors. These pathways operate mainly by receiving signals from growth factors that bind to cell-surface receptors typically with intracellular tyrosine kinase activity [72]. This activity modulates pro-survival signaling pathways such as phosphatidylinositol 3-kinase/protein kinase B (PI3K/AKT), the mammalian target of rapamycin (mTOR) [73] and the mitogen-activated protein kinases/extracellular signal-regulated kinase (MAPK/ERK) [74]. The development of specific receptor tyrosine kinase inhibitors has resulted in effective chemotherapeutics [74]. However, cancer cells can also utilize many other pathways to proliferate and eventually circumvent the inhibited routes in order to develop resistance [75, 76]. Repurposed drugs may provide a solution to this problem. For example, rapamycin, an mTOR1 inhibitor initially approved as an immunosuppressant and later as an anti-restenosis agent [77, 78], was repurposed to treat leukemias due to the importance of the mTOR pathway in cancer [79, 80] [81]. However, inhibition of mTOR1 is often compensated by the activation of PI3K-AKT [82] and the reactivation of eukaryotic translation initiation factor 4E-binding proteins [83, 84]. Despite these limitations, rapamycin and its more soluble and specific analogs, like temsirolimus [85], have been tested in combination with growth factor receptor antagonists [86].

Other candidates for repurposing that could target cancer proliferation include prazosin, an alpha blocker initially approved to treat hypertension [87], and indomethacin, a non-steroidal anti-inflammatory drug (NSAID) [88]. Prazosin has been reported to inhibit AKT signaling [89, 90] and is recommended for treating pheochromocytoma [91]. It is also included in a Phase 1 study as an additive to radiotherapy in men with prostate cancer (ACTRN12621000784819). Indomethacin, besides its Cox1/2 inhibition-related antiangiogenic effects [92], has been shown to impair cancer proliferation by inhibiting MAPK [93] or PKC signaling [94]. Currently, three indomethacin-related clinical trials are registered, including a Phase 4 trial for prostate cancer (ChiCTR2000038968) and a Phase 1 study for breast cancer (NCT02950259). Furthermore, drugs that could effectively target human telomerase reverse transcriptase (hTERT) in cancer cells to limit their replication remain an interest for drug repurposing strategies [95].

Another promising strategy is the search for drugs that can accelerate cancer cell death. Along with progress in understanding the molecular mechanisms underlying regulated cell death [96, 97], many related adverse effects of non-oncological drugs may be useful in cancer therapies. Considering that cancer cells often circumvent common apoptotic pathways, drugs that can selectively accelerate other cell death modalities— including ferroptosis [98], cuproptosis [99], necroptosis [100], pyroptosis[101], and lethal autophagy [102]—can improve treatment strategies while reducing general toxicity. One exception, however, is autophagy since this can also favor cancer cells by enhancing their survival under metabolic and environmental stresses [103], and therefore should be considered with caution.

Artemisinin and chloroquine, along with their derivatives were initially dedicated for treating malaria [104], are well-known examples of cell death-related drug repurposing. Artemisinins have been reported to induce non-apoptotic programmed cell death, especially ferroptosis, in cancer cells [105–109]. Recent clinical trials for artesunate are testing its application in colorectal cancers (NCT02633098) and leukemias (CTRI/2024/03/063617). Chloroquine and its derivative hydroxychloroquine are approved as autophagy flux inhibitors to treat pancreatic and other cancers [110, 111]. Currently, these compounds are undergoing 48 clinical trials related to cancer therapies (Database Repurposing Trials In Oncology, ReDO_Trials_DB, https://www.anticancer fund.org/en/database-repurposing-trials-oncology as for 14.08.2024) [112].

Mebendazole (5-benzoyl-1H-benzimidazol-2-ylcarbamate), initially intended as an anthelmintic agent [113], is another candidate for repurposing into oncological therapies due to its potential to inhibit microtubule polymerization [114, 115]. Notably, besides restricting tumor growth, this compound has been effective in preventing the invasion and metastasis of malignant tumors and glioblastomas [116, 117]. Zhang [118] and in other individual cancer cases [114, 115]. Currently, six mebendazole-related clinical trials are registered in the ReDO_Trials_DB [112].

Deregulated metabolism and increased energy demands are other hallmarks of cancer [119, 120] that have been targeted for drug repurposing [121]. Patients with diabetes are generally more prone to several types of cancer [122, 123]. Long-term treatment with metformin, approved for obese type 2 diabetes [124], has been observed to lower the risk of cancer in diabetic patients, making this drug a potential candidate for repurposing [120, 122]. Indeed, there are 133 metformin-related trials in the ReDO_Trials_DB [112]. However, despite these efforts, results have been far from satisfactory [125, 126]. Furthermore, the mechanism of action of metformin in tumor cells and the tumor microenvironment remains unclear and under discussion. Although high doses of this compound impair cellular respiration by inhibiting Complex-1 [127], preclinical studies have observed a plethora of pleiotropic effects of metformin administration in cancer cells that are independent of Complex-1 inhibition [128, 129]. Hopefully, continuous research on this compound will eventually allow successful metformin repurposing.

Disulfiram provides another example of a repurposed drug, initially intended to treat alcoholism [130], now targeting cancer cell metabolism ^{129,130}. Disulfiram inhibits acetaldehyde dehydrogenase activity [131], resulting in alcohol intolerance, as well as the blockage of formaldehyde oxidation [132] and deregulated oxidative metabolism in cancer cells [133]. Furthermore, the p97 segregase adaptor NPL4 (also known as VCP), important for maintaining cellular proteostasis, has been identified as the molecular target of disulfiram responsible for its anticancer activity [134]. Currently, there are 13 disulfiram clinical trials in the ReDO_Trials_DB, including a phase 2/3 application of this drug for glioblastoma (NCT02678975) [112].

The ability of cancers to impair and circumvent host immune responses provides another therapeutic opportunity [135–138]. Pharmaceutical solutions that could increase antitumor immunity, such as immune checkpoint inhibitors, are currently of great interest [139]. It has been observed that some vaccines dedicated to infectious diseases (such as rotaviruses, yellow fever, and influenza), when administered intratumorally, can activate antitumor immunity [140–143].

Other drug repurposing strategies arise from the specific impairment of tumor suppressors such as p53 or the retinoblastoma protein in cancer cells [144]. Along these lines, quinacrine, an antimalarial agent [145], was found to be a promising candidate since it was reported to induce p53 expression in cancer cells [146] and can exert some anticancer activity in a p53-dependent manner [146, 147]. The accumulation of p53 has also been observed in cancer cells treated with ritonavir, a protease inhibitor used to treat human immunodeficiency virus (HIV) infection [148, 149]. Furthermore, other reports found this compound capable of reactivating the retinoblastoma protein [147]. Currently, there is one active phase 1 clinical trial for the application of ritonavir for prostate cancer (NCT05679388) [112]. Notably, statins have also been shown to increase p53 activity in cancer cells and thus display anticancer potential [148, 149]. Currently, there are 47 statin-related records in the ReDO_Trials_DB, many of which are reaching phase 3 or 4 [112].

Tumor expansion is accompanied by an increased demand for nutrients and oxygen by cancer cells, leading to the induction of chronic angiogenesis [150]. Although this hall-mark of cancer cells has resulted in the development of currently used antiangiogenic agents that limit tumor blood flow and lead to its starvation [58, 151], the use of these drugs is limited and can unfortunately stimulate resistance [152]. Notably, the repurposing of thalidomide, an immunomodulatory drug initially sold to treat morning sickness that was withdrawn worldwide in 1962 after it was linked to severe birth defects [153], has shown promise. Thalidomide still remains in use to treat leprosy [153] and is currently used as an antiangiogenic agent [154, 155]. It is also approved for combination therapy in multiple myeloma [156].

Antiangiogenic and anticancer potential has also been observed for the approved antifungal agent itraconazole [157, 158]. Currently, there are 17 itraconazole-related records in the ReDO_Trials_DB, one of which for ovarian cancer is reaching phase 3 (NCT03458221). Interestingly, artemisinins have also been assigned antiangiogenic activities [159]. In contrast, it has been reported that some anticancer approaches aim to induce angiogenesis in order to facilitate drugs delivery [160, 161].

Notably, cancer-related inflammation supports not only angiogenesis but also invasion and metastasis, as well as reprogramming of the tumor microenvironment (TME) [162, 163]. Therefore, anti-inflammatory drugs could be good candidates for repurposing. Celecoxib, a selective cyclooxygenase-2 (COX-2) inhibitor approved for adult arthritis [164], is extensively being tested in clinical trials (57 registered in ReDO_Trials_DB) due to multiple reports of its potential to enhance the chemosensitivity of cancer cells and to reduce the toxicity of marketed chemotherapeutics [165–167]. Similarly, 31 clinical trials are registered for aspirin (ReDO Trials DB), which was already suggested to have potential as an anticancer drug in the 1970s [168]. More recent research reports of aspirin being effective against many types of cancer [169–171] are supported by epidemiological analyses [172-175]. Indeed, low-dose aspirin inhibits the production of thromboxane A2 (TXA2) by irreversibly inhibiting the enzyme COX-1 in platelets. By reducing TXA2, aspirin can help prevent the formation of blood clots and has been shown to have potential benefits in reducing the risk of cancer progression and metastasis [176, 177]. Furthermore, aspirin at higher doses is also a more potent COX-2 inhibitor, which can increase its anti-cancer properties against tumors that overexpress this enzyme [175]. Numerous studies shown that daily low-doses of aspirin may significantly reduce the risk of colon cancer and rectal cancer [178] and breast cancer [179, 180]. Recent results from a 20-year cohort study involving 1,909,531 individuals in Denmark have shown that long-term low-dose aspirin use is associated with slightly to moderately reduced risks for several specific cancers [181]. However, there was no reduction in overall cancer risk for some common cancers [181]. Similar or slightly stronger inverse associations were observed for the consistent use of high-dose aspirin [181].

Interestingly, the ASPirin in Reducing Events in the Elderly (ASPREE) study, which was double-blind and performed on a large cohort (for 4.7 years and over 19,000 individuals older than 65–70, that did not have cardiovascular disease, dementia, or disability), showed no advantage of taking low-dose aspirin, and in fact increased the risk of being diagnosed with stage 3 or 4 cancers as well as increased mortality rates compared to the placebo [182]. In contrast, prior randomized controlled trials, mainly involving younger individuals, demonstrated a delayed cancer benefit with aspirin [182]. However, these study conclusions are under discussion and should be taken with caution, as deaths were classified according to the underlying cause by adjudicators who were unaware of the trial-group assignments. Furthermore, hazard ratios were calculated to compare mortal-ity between the aspirin group and the placebo group, and post hoc exploratory analyses of specific causes of death were performed [182].

Furthermore, some of the adverse effects of many approved chemotherapeutics can be reduced with the use of additional anti-inflammatory compounds and beta-blockers to reduce cardiotoxicity [183, 184]. The latter (especially propranolol and timolol) have been reported to have anticancer activity [185–187]. Taken together, these examples provided illustrate the vast potential of drug repurposing in oncology.

Discussion

Although the drug repurposing approach seems like an attractive solution to benefit both cancer patients and health systems, its results have been below expectations despite the considerable efforts and high academic attention. Continuous research and technological development have yet to yield breakthrough solutions. While there are some examples of successful drug repositioning, the majority of candidates, like metformin, have remained in the reassignment pipelines for many years. As of August 14, 2024, the ReDO_Trials_DB database reports 898 (409 controlled) trials related to 182 drugs (with metformin being the most commonly tested one) and involving 157,295 patients [112] (Fig. 1). Notably, however, less than 5% of these trials are sponsored by

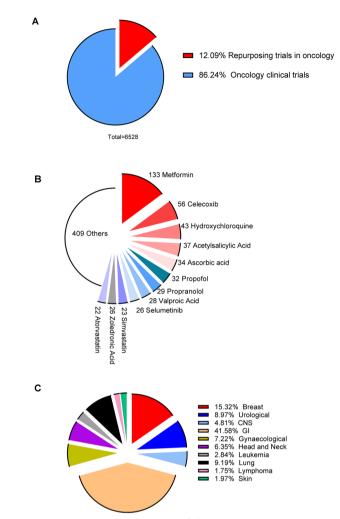


Fig. 1 A. Repurposing trials are a modest percentage of all oncology clinical trials. **B** The most popular drugs in repurposing trials (**C**) and the most popular cancers to be targeted in controlled trails of repurposed drugs. The data were obtained from the ReDO_Trials_DB database and the GlobalData's Clinical Trials Intelligence Center (December 15, 2023)

pharmaceutical companies, with the vast majority funded by non-profit organizations [188, 189]. Furthermore, 139 clinical trials have reached phase 3 or 4 [112].

Although these numbers may initially seem impressive, they are modest compared to data from the GlobalData's Clinical Trials Intelligence Center ((https://www.clinicaltrialsarena.com/sponsored/oncology-in-2024-the-clinical-trial-trends-reshaping-the-role-of-cros/). As of December 15, 2023, 6,528 oncology-related trials were noted, with another 569 planned to begin. In the DrugBank database ((https://go.drugbank.com/releases/latest), there are 4,493 approved drugs (2812 small molecules and 1681 biotechnology drugs) that can be directed against over 3,000 unique targets (Fig. 1). Additionally, there are over 333 withdrawn drugs (302 small molecules and 31 biotechnology drugs), 8,000 investigational ones (5311 small molecules and 2752 biotechnology drugs), and about 6732 experimental compounds (6353 small molecules and 379 biotechnology drugs). Taken together, drug repurposing in oncology, despite its potential and large

number of possible candidates, remains underappreciated and mostly limited to academic research and small biotech companies.

Notably, the business models of Big Pharma rely strongly on market exclusivity for their drugs, allowing them to sell their drugs at high prices [190–192]. Since repurposed drugs cannot usually be considered novel chemical entities and their structures are already known, novel patent claims to the active pharmaceutical ingredient are not possible [193]. The repurposed drug can only be protected at the level of the 'method of use' for the new indication, although such protection is harder to obtain and costs more [193]. Finally, the use of patents excludes off-label prescriptions, where medications are prescribed for indications or populations for which they have no regulatory approval [194]. Numerous solutions have been suggested to motivate Big Pharma efforts towards repurposing (tax breaks, FDA-priority review vouchers, or funding clinical trials) [195, 196]. However, given the current business model Big Pharma operates under, implementing these initiatives probably will not be a game changer [188]. This conflict of interest is well illustrated by thalidomide approvals. Despite this drug in combination with melphalan–prednisone is comparable to the dramatically more expensive lenalido-mide, the more expensive new drug was approved as the standard therapy [197, 198].

Indeed, mainly academic and independent research provides the rationale for using off-patent medications in cancer treatment [40]. However, in the case of drug repurposing, these academic approaches often lack specific insights that are exclusive to pharmaceutical companies. Due to limited resources, lack of data, technology, funding, and experience, many academic attempts at drug repurposing are often "fashion" driven (metformin, statins, aspirin, ascorbic acid, etc.) [112] and thus oriented on drugs that may be easy to obtain, publish, and get funded for their application (Fig. 1B). However, this approach does not consider that millions of people who are cancer patients or will develop cancers are already taking these prescriptions, whereas their benefits on a wide population scale in terms of cancer risks are usually under discussion or not properly documented.

Furthermore, academic research is the main contributor of related omics and epidemiological data that are further used for machine learning and other applications utilized by drug repositioning strategies. Notably, retrospective observational studies are subject to immortal time bias and selection bias [199], resulting in frequent overestimations of their advantages for the treatment group [200]. This case is well illustrated by the numerous correlations between metformin treatments and the incidence of cancer [201]. Furthermore, since metformin's original target group were diabetic patients who differ from cancer patients, selection bias occurred [202]. Additionally, computational strategies for predicting drug reassignment are only as good as the reliability of the input data and can often discourage target-centric and drug-centric strategies. Phenotypebased high throughput screening strategies, due to the costs of libraries and extensive labor, are usually beyond basic research funding schemes. Finally, the costs and ability to design dedicated and sufficient clinical trials that can include various cancer types and patient groups, remain another serious barrier for academic approaches.

The complexity of cancer limits the efficacy of monotherapies, whereas combination therapies come with specific challenges and limitations that often translate into drug repurposing approaches [199, 203, 204]. Notably, combination therapy trials are more

complex and thus cost more than those for monotherapies [205]. Since phase 2 trials are usually the turning point for a drug's fate, progressing them towards more randomized larger cohort research trials requires promising efficacy results in order to justify the financial burden [199].

Many clinical trials fail due to insufficient cancer patient accrual [206]. Cancer patients are often older people with accompanying diseases and undergoing different cancerrelated or unrelated treatments, which may result in unexpected side effects and death during the trials [33, 207]. Indeed, combinational therapies do not guarantee better efficacy [33]. For example, combined dacarbazine with cisplatin, carmustine, and tamoxifen for metastatic (stage IV) melanoma treatment is comparable in terms of patient survival with monotherapy based on high-dose dacarbazine [207, 208]. Unfortunately, dosages that may be well tolerated in trials conducted using healthy subjects or when used for treating the diseases the drug was originally intended for may not be achievable in cancer patients [33]. Repurposed drugs intended for specific targets might not have the same efficacy in cancer cells or in the presence of other drugs [33]. Therefore, their repositioning may require higher doses, which can result in novel distinct mechanisms of action and consequently unforeseen adverse effects [209, 210]. For example, aspirin repurposed for use in high doses may lead to an increased risk of gastrointestinal bleeding, while simvastatin and metformin contribute to the development of hypolipidemia and hypoglycemia. All these issues stress the importance of quality preclinical research that will allow only the most effective and safest combinations to be trialed in phases 2 and 3 [204, 211, 212].

Importantly, recent scientific and technological breakthroughs are becoming more economically available and thus possible to incorporate into high-throughput screening pipelines, as well as drug-centric approaches. Starting with omics approaches, which allow the determination of molecular mechanisms of repurposing candidates, and coupling this with failed drugs could provide valuable insight into the cellular proteomic, metabolomic, and transcriptomic changes [44, 213–222] in various cancers. Importantly, single-cell sequencing seems to be a way to address cancer heterogeneity [223], while the rapid growth and development of databases provide machine learning and computational approaches with more reliable insights [224–227]. Furthermore, the development of novel drug delivery methods that allow more specific and even compartment-targeted application of repurposed drugs may contribute to the success of novel repurposing strategies [228–230].

Conclusions and perspectives

Repurposed drugs have the undeniable potential to address the shortage of new drugs and combat acquired chemotherapy resistance. Additionally, many healthcare systems struggle to provide patients with expensive new generations of chemotherapeutics, let alone personalized therapies. The financial advantages of drug repositioning could benefit many patients worldwide. However, despite research progress, multiple pharmacological challenges and obstacles need to be addressed to effectively utilize the opportunity of repurposed drugs in cancer treatment. Importantly, the field requires programs, regulations, and government-level funding to promote and support collaboration between academia and the pharmaceutical industry [40, 190]. Such programs in the USA and UK have allowed the transfer of a large number of failed drugs to academic research for repurposing [231]. Furthermore, continuous efforts and lobbying by academic and non-profit organizations are necessary to popularize and develop new drug discovery concepts.

Abbreviations

AKT ASPREE CLI	Protein kinase B ASPirin in Reducing Events in the Elderly Chronic lymphocytic leukemia
CML	Chronic myeloid leukemia
COX-2	Cyclooxygenase-2
ERK	Extracellular signal-regulated kinase
FDA	Food and Drug Administration
GWASs	Genome-wide association studies
HIV	Human immunodeficiency virus
htert	Human telomerase reverse transcriptase
MAPK	Mitogen-activated protein kinases
mTOR	Mammalian target of rapamycin
NSAID	Non-steroidal anti-inflammatory drug
PI3K	Phosphatidylinositol 3-kinase
R&D	Research and development
TME	Tumor microenvironment

Acknowledgements

We would like to apologize to all those who have made significant and important contributions to our understanding of this research area that are not referenced in this manuscript.

Author contributions

All authors wrote and revised the paper.

Funding

Not applicable.

Availability of data and materials Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing financial interests.

Received: 29 November 2024 Accepted: 24 February 2025 Published online: 05 March 2025

References

- Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, Jemal A. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA: A Cancer J Clin. 2024;74(3):229–63.
- 2. Global Burden of Disease Cancer C. The global burden of cancer 2013. JAMA Oncol. 2015;1(4):505–27.
- Holohan C, Van Schaeybroeck S, Longley DB, Johnston PG. Cancer drug resistance: an evolving paradigm. Nat Rev Cancer. 2013;13(10):714–26.
- 4. Anderson NM, Simon MC. The tumor microenvironment. Curr Biol. 2020;30(16):R921-5.
- 5. Baghban R, Roshangar L, Jahanban-Esfahlan R, Seidi K, Ebrahimi-Kalan A, Jaymand M, et al. Tumor
- microenvironment complexity and therapeutic implications at a glance. Cell Communi Signal. 2020;18(1):59.
 de Visser KE, Joyce JA. The evolving tumor microenvironment: from cancer initiation to metastatic outgrowth. Cancer Cell. 2023;41(3):374–403
- Bartoszewska S, Collawn JF, Bartoszewski R. The role of the hypoxia-related unfolded protein response (UPR) in the tumor microenvironment. Cancers. 2022;14(19):4870.

- 8. Qin S-Y, Cheng Y-J, Lei Q, Zhang A-Q, Zhang X-Z. Combinational strategy for high-performance cancer chemotherapy. Biomaterials. 2018;171:178–97.
- Vokinger KN, Hwang TJ, Grischott T, Reichert S, Tibau A, Rosemann T, Kesselheim AS. Prices and clinical benefit of cancer drugs in the USA and Europe: a cost-benefit analysis. Lancet Oncol. 2020;21(5):664–70.
- 10. Sun D, Gao W, Hu H, Zhou S. Why 90% of clinical drug development fails and how to improve it? Acta Pharm Sin B. 2022;12(7):3049–62.
- 11. Scannell JW, Blanckley A, Boldon H, Warrington B. Diagnosing the decline in pharmaceutical R&D efficiency. Nat Rev Drug Discov. 2012;11(3):191–200.
- 12. Hay M, Thomas DW, Craighead JL, Economides C, Rosenthal J. Clinical development success rates for investigational drugs. Nat Biotechnol. 2014;32(1):40–51.
- 13. Milsted RA. Cancer drug approval in the United States, Europe, and Japan. Adv Cancer Res. 2007;96:371-91.
- 14. Bateman-House A, Robertson CT. The federal right to try act of 2017-A wrong turn for access to investigational drugs and the path forward. JAMA Intern Med. 2018;178(3):321–2.
- Khozin S, Liu K, Jarow JP, Pazdur R. Regulatory watch: why do oncology drugs fail to gain US regulatory approval? Nat Rev Drug Discov. 2015;14(7):450–1.
- 16. Benderly BL. Experimental drugs on trial. Sci Am. 2007;297(4):92-9.
- 17. FDA treads delicate line between safety and speed. Oncology (Williston Park). 1999;13(1):16
- Iazzolino G, Bozzo R. Partnership models for R & D in the pharmaceutical industry. In: Canci JK, Mekler P, Mu G, editors. Quantitative models in life science business: from value creation to business processes. Cham: Springer International Publishing; 2023. p. 29–48.
- 19. Bowes J, Brown AJ, Hamon J, Jarolimek W, Sridhar A, Waldron G, Whitebread S. Reducing safety-related drug attrition: the use of in vitro pharmacological profiling. Nat Rev Drug Discov. 2012;11(12):909–22.
- 20. Kang H, Pan S, Lin S, Wang Y-Y, Yuan N, Jia P. PharmGWAS: a GWAS-based knowledgebase for drug repurposing. Nucleic Acids Res. 2024;52(D1):D972–9.
- 21. Ringel MS, Scannell JW, Baedeker M, Schulze U. Breaking Eroom's Law. Nat Rev Drug Discov. 2020;19(12):833-4.
- 22. Pushpakom S, Iorio F, Eyers PA, Escott KJ, Hopper S, Wells A, et al. Drug repurposing: progress, challenges and recommendations. Nat Rev Drug Discov. 2019;18(1):41–58.
- Parisi D, Adasme MF, Sveshnikova A, Bolz SN, Moreau Y, Schroeder M. Drug repositioning or target repositioning: a structural perspective of drug-target-indication relationship for available repurposed drugs. Comput Struct Biotechnol J. 2020;18:1043–55.
- Kirtonia A, Gala K, Fernandes SG, Pandya G, Pandey AK, Sethi G, et al. Repurposing of drugs: an attractive pharmacological strategy for cancer therapeutics. Semin Cancer Biol. 2021;68:258–78.
- Huang A, Garraway LA, Ashworth A, Weber B. Synthetic lethality as an engine for cancer drug target discovery. Nat Rev Drug Discov. 2020;19(1):23–38.
- Dallavalle S, Dobričić V, Lazzarato L, Gazzano E, Machuqueiro M, Pajeva I, et al. Improvement of conventional anticancer drugs as new tools against multidrug resistant tumors. Drug Resist Updat. 2020;50:100682.
- Patel MN, Halling-Brown MD, Tym JE, Workman P, Al-Lazikani B. Objective assessment of cancer genes for drug discovery. Nat Rev Drug Discov. 2013;12(1):35–50.
- 28. Eder J, Sedrani R, Wiesmann C. The discovery of first-in-class drugs: origins and evolution. Nat Rev Drug Discov. 2014;13(8):577–87.
- Swinney DC. Phenotypic vs. target-based drug discovery for first-in-class medicines. Clin Pharmacol Ther. 2013;93(4):299–301.
- Moffat JG, Vincent F, Lee JA, Eder J, Prunotto M. Opportunities and challenges in phenotypic drug discovery: an industry perspective. Nat Rev Drug Discov. 2017;16(8):531–43.
- 31. Adair FE, Bagg HJ. Experimental and clinical studies on the treatment of cancer by dichlorethylsulphide (mustard gas. Ann Surg. 1931;93(1):190–9.
- 32. Haddow A. On the biological alkylating agents. Perspect Biol Med. 1973;16(4):503-24.
- 33. Schein CH. Repurposing approved drugs for cancer therapy. Br Med Bull. 2021;137(1):13–27.
- Maxmen A. Busting the billion-dollar myth: how to slash the cost of drug development. Nature. 2016;536(7617):388–90.
- 35. Bertolini F, Sukhatme VP, Bouche G. Drug repurposing in oncology–patient and health systems opportunities. Nat Rev Clin Oncol. 2015;12(12):732–42.
- 36. Mullard A. Biotech R&D spend jumps by more than 15. Nat Rev Drug Discov. 2016;15(7):447.
- 37. Weir SJ, DeGennaro LJ, Austin CP. Repurposing approved and abandoned drugs for the treatment and prevention of cancer through public-private partnership. Cancer Res. 2012;72(5):1055–8.
- Fogel DB. Factors associated with clinical trials that fail and opportunities for improving the likelihood of success: a review. Contemp Clin Trials Commun. 2018;11:156–64.
- 39. Cha Y, Erez T, Reynolds IJ, Kumar D, Ross J, Koytiger G, et al. Drug repurposing from the perspective of pharmaceutical companies. Br J Pharmacol. 2018;175(2):168–80.
- 40. Weth FR, Hoggarth GB, Weth AF, Paterson E, White MPJ, Tan ST, et al. Unlocking hidden potential: advancements, approaches, and obstacles in repurposing drugs for cancer therapy. Br J Cancer. 2024;130(5):703–15.
- 41. Moffat JG, Vincent F, Lee JA, Eder J, Prunotto M. Opportunities and challenges in phenotypic drug discovery: an industry perspective. Nat Rev Drug Discov. 2017;16(8):531–43.
- 42. Jordan VC. Tamoxifen: a most unlikely pioneering medicine. Nat Rev Drug Discov. 2003;2(3):205-13.
- 43. Pessetto ZY, Weir SJ, Sethi G, Broward MA, Godwin AK. Drug repurposing for gastrointestinal stromal tumor. Mol Cancer Ther. 2013;12(7):1299–309.
- Macioszek S, Dudzik D, Bartoszewski R, Stokowy T, Lambrechts D, Boeckx B, et al. Metabolomic and transcriptomic response to imatinib treatment of gastrointestinal stromal tumour in xenograft-bearing mice. Transl Oncol. 2023;30:101632.
- 45. Huang X, Guo B. Adenomatous polyposis coli determines sensitivity to histone deacetylase inhibitor-induced apoptosis in colon cancer cells. Can Res. 2006;66(18):9245–51.

- Meng XY, Zhang HX, Mezei M, Cui M. Molecular docking: a powerful approach for structure-based drug discovery. Curr Comput Aided Drug Des. 2011;7(2):146–57.
- Giordano D, Biancaniello C, Argenio MA, Facchiano A. Drug design by pharmacophore and virtual screening approach. Pharmaceuticals. 2022. https://doi.org/10.3390/ph15050646.
- Salentin S, Schreiber S, Haupt VJ, Adasme MF, Schroeder M. PLIP: fully automated protein–ligand interaction profiler. Nucleic Acids Res. 2015;43(W1):W443–7.
- 49. Yan X-Y, Zhang S-W, Zhang S-Y. Prediction of drug–target interaction by label propagation with mutual interaction information derived from heterogeneous network. Mol BioSyst. 2016;12(2):520–31.
- 50. Catacutan DB, Alexander J, Arnold A, Stokes JM. Machine learning in preclinical drug discovery. Nat Chem Biol. 2024;20(8):960–73.
- Pahikkala T, Airola A, Pietilä S, Shakyawar S, Szwajda A, Tang J, Aittokallio T. Toward more realistic drug-target interaction predictions. Brief Bioinform. 2015;16(2):325–37.
- 52. Gore SD. New ways to use DNA methyltransferase inhibitors for the treatment of myelodysplastic syndrome. Hematol Am Soc Hematol Educ Program. 2011;2011:550–5.
- 53. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646-74.
- 54. Hanahan D. Hallmarks of cancer: new dimensions. Cancer Discov. 2022;12(1):31-46.
- Flavahan WA, Gaskell E, Bernstein BE. Epigenetic plasticity and the hallmarks of cancer. Science. 2017. https://doi. org/10.1126/science.aal2380.
- Sun W, Sanderson PE, Zheng W. Drug combination therapy increases successful drug repositioning. Drug Discov Today. 2016;21(7):1189–95.
- Boshuizen J, Peeper DS. Rational cancer treatment combinations: an urgent clinical need. Mol Cell. 2020;78(6):1002–18.
- Slawski J, Jaśkiewicz M, Barton A, Kozioł S, Collawn JF, Bartoszewski R. Regulation of the HIF switch in human endothelial and cancer cells. Eur J Cell Biol. 2024;103(2):151386.
- Białek W, Hryniewicz-Jankowska A, Czechowicz P, Sławski J, Collawn JF, Czogalla A, Bartoszewski R. The lipid side of unfolded protein response. Biochim et Biophys Acta (BBA) – Mol Cell Biol Lip. 2024;1869(7):159515.
- 60. Bartoszewska S, Cabaj A, Dąbrowski M, Collawn JF, Bartoszewski R. miR-34c-5p modulates X-box-binding protein 1 (XBP1) expression during the adaptive phase of the unfolded protein response. Faseb J. 2019;33(10):11541–54.
- Bartoszewski R, Gebert M, Janaszak-Jasiecka A, Cabaj A, Króliczewski J, Bartoszewska S, et al. Genome-wide mRNA
 profiling identifies RCAN1 and GADD45A as regulators of the transitional switch from survival to apoptosis during
 ER stress. FEBS J. 2020;287(14):2923–47.
- Gebert M, Sobolewska A, Bartoszewska S, Cabaj A, Crossman DK, Króliczewski J, et al. Genome-wide mRNA profiling identifies X-box-binding protein 1 (XBP1) as an IRE1 and PUMA repressor. Cell Mol Life Sci. 2021;78(21):7061–80.
- 63. Payne KK. Cellular stress responses and metabolic reprogramming in cancer progression and dormancy. Semin Cancer Biol. 2022;78:45–8.
- Aranda-Anzaldo A, Dent MAR. Is cancer a disease set up by cellular stress responses? Cell Stress Chaperones. 2021;26(4):597–609.
- 65. Fulda S, Gorman AM, Hori O, Samali A. Cellular stress responses: cell survival and cell death. Int J Cell Biol. 2010;2010(1):214074.
- 66. Moszyńska A, Collawn JF, Bartoszewski R. IRE1 endoribonuclease activity modulates hypoxic HIF-1α signaling in human endothelial cells. Biomolecules. 2020;10(6):895.
- Moszyńska A, Jaśkiewicz M, Serocki M, Cabaj A, Crossman DK, Bartoszewska S, et al. The hypoxia-induced changes in miRNA-mRNA in RNA-induced silencing complexes and HIF-2 induced miRNAs in human endothelial cells. Faseb J. 2022;36(7):e22412.
- Bartoszewska S, Sławski J, Collawn JF, Bartoszewski R. Dual RNase activity of IRE1 as a target for anticancer therapies. J Cell Communi Signal. 2023;17(4):1145–61.
- 69. Gebert M, Bartoszewska S, Opalinski L, Collawn JF, Bartoszewski R. IRE1-mediated degradation of pre-miR-301a promotes apoptosis through upregulation of GADD45A. Cell Commun Signal. 2023;21(1):322.
- Gebert M, Sławski J, Kalinowski L, Collawn JF, Bartoszewski R. The unfolded protein response: a double-edged sword for brain health. Antioxidants. 2023;12(8):1648.
- 71. Tiligada E. Chemotherapy: induction of stress responses. Endocr Relat Cancer. 2006;13(Suppl 1):S115–24.
- 72. Du Z, Lovly CM. Mechanisms of receptor tyrosine kinase activation in cancer. Mol Cancer. 2018;17(1):58.
- 73. Panwar V, Singh A, Bhatt M, Tonk RK, Azizov S, Raza AS, et al. Multifaceted role of mTOR (mammalian target of rapamycin) signaling pathway in human health and disease. Signal Transduct Target Ther. 2023;8(1):375.
- Pottier C, Fresnais M, Gilon M, Jérusalem G, Longuespée R, Sounni NE. Tyrosine kinase inhibitors in cancer: breakthrough and challenges of targeted therapy. Cancers. 2020. https://doi.org/10.3390/cancers12030731.
- Rodrik-Outmezguine VS, Chandarlapaty S, Pagano NC, Poulikakos PI, Scaltriti M, Moskatel E, et al. mTOR kinase inhibition causes feedback-dependent biphasic regulation of AKT signaling. Cancer Discov. 2011;1(3):248–59.
- Ahronian LG, Sennott EM, Van Allen EM, Wagle N, Kwak EL, Faris JE, et al. Clinical acquired resistance to RAF inhibitor combinations in BRAF-mutant colorectal cancer through MAPK pathway alterations. Cancer Discov. 2015;5(4):358–67.
- 77. Benjamin D, Colombi M, Moroni C, Hall MN. Rapamycin passes the torch: a new generation of mTOR inhibitors. Nat Rev Drug Discov. 2011;10(11):868–80.
- Farb A, John M, Acampado E, Kolodgie FD, Prescott MF, Virmani R. Oral everolimus inhibits in-stent neointimal growth. Circulation. 2002;106(18):2379–84.
- 79. Grabiner BC, Nardi V, Birsoy K, Possemato R, Shen K, Sinha S, et al. A diverse array of cancer-associated MTOR mutations are hyperactivating and can predict rapamycin sensitivity. Cancer Discov. 2014;4(5):554–63.
- Altman JK, Sassano A, Kaur S, Glaser H, Kroczynska B, Redig AJ, et al. Dual mTORC2/mTORC1 targeting results in potent suppressive effects on acute myeloid leukemia (AML) progenitors. Clin Cancer Res. 2011;17(13):4378–88.

- 81. Sillaber C, Mayerhofer M, Böhm A, Vales A, Gruze A, Aichberger K, et al. Evaluation of antileukaemic effects of rapamycin in patients with imatinib-resistant chronic myeloid leukaemia. Eur J Clin Invest. 2008;38(1):43–52.
- O'Reilly KE, Rojo F, She Q-B, Solit D, Mills GB, Smith D, et al. mTOR inhibition induces upstream receptor tyrosine kinase signaling and activates Akt. Can Res. 2006;66(3):1500–8.
- Choo AY, Yoon SO, Kim SG, Roux PP, Blenis J. Rapamycin differentially inhibits S6Ks and 4E-BP1 to mediate celltype-specific repression of mRNA translation. Proc Natl Acad Sci U S A. 2008;105(45):17414–9.
- Dowling RJ, Topisirovic I, Alain T, Bidinosti M, Fonseca BD, Petroulakis E, et al. mTORC1-mediated cell proliferation, but not cell growth, controlled by the 4E-BPs. Science. 2010;328(5982):1172–6.
- Malizzia LJ, Hsu A. Temsirolimus, an mTOR inhibitor for treatment of patients with advanced renal cell carcinoma. Clin J Oncol Nurs. 2008;12(4):639–46.
- Rugo HS, Trédan O, Ro J, Morales SM, Campone M, Musolino A, et al. A randomized phase II trial of ridaforolimus, dalotuzumab, and exemestane compared with ridaforolimus and exemestane in patients with advanced breast cancer. Breast Cancer Res Treat. 2017;165:601–9.
- Kirkendall WM, Hammond JJ, Thomas JC, Overturf ML, Zama A. Prazosin and clonidine for moderately severe hypertension. JAMA. 1978;240(23):2553–6.
- Percy J, Stephenson P, Thompson M. Indomethacin in the treatment of rheumatic diseases. Ann Rheum Dis. 1964;23(3):226.
- Desiniotis A, Kyprianou N. Advances in the design and synthesis of prazosin derivatives over the last ten years. Expert Opin Ther Targets. 2011;15(12):1405–18.
- Assad Kahn S, Costa SL, Gholamin S, Nitta RT, Dubois LG, Fève M, et al. The anti-hypertensive drug prazosin inhibits glioblastoma growth via the PKC δ-dependent inhibition of the AKT pathway. EMBO Mol Med. 2016;8(5):511–26.
- 91. Nicholson JP Jr, Vaughn ED Jr, Pickering TG, Resnick LM, Artusio J, Kleinert HD, et al. Pheochromocytoma and prazosin. Ann Internal Med. 1983;99(4):477–9.
- Touhey S, O'Connor R, Plunkett S, Maguire A, Clynes M. Structure–activity relationship of indomethacin analogues for MRP-1, COX-1 and COX-2 inhibition: identification of novel chemotherapeutic drug resistance modulators. Eur J Cancer. 2002;38(12):1661–70.
- 93. Lin C-C, Suen KM, Stainthorp A, Wieteska L, Biggs GS, Leitão A, et al. Targeting the Shc-EGFR interaction with indomethacin inhibits MAP kinase pathway signalling. Cancer Lett. 2019;457:86–97.
- Mazumder S, De R, Debsharma S, Bindu S, Maity P, Sarkar S, et al. Indomethacin impairs mitochondrial dynamics by activating the PKCζ–p38–DRP1 pathway and inducing apoptosis in gastric cancer and normal mucosal cells. J Biol Chem. 2019;294(20):8238–58.
- Vishwakarma K, Dey R, Bhatt H. Telomerase: a prominent oncological target for development of chemotherapeutic agents. Eur J Med Chem. 2023;249:115121.
- 96. Galluzzi L, Vitale I, Aaronson SA, Abrams JM, Adam D, Agostinis P, et al. Molecular mechanisms of cell death: recommendations of the nomenclature committee on cell death 2018. Cell Death Differ. 2018;25(3):486–541.
- 97. Peng F, Liao M, Qin R, Zhu S, Peng C, Fu L, et al. Regulated cell death (RCD) in cancer: key pathways and targeted therapies. Signal Transduct Target Ther. 2022;7(1):286.
- Dixon SJ, Lemberg KM, Lamprecht MR, Skouta R, Zaitsev EM, Gleason CE, et al. Ferroptosis: an iron-dependent form of nonapoptotic cell death. Cell. 2012;149(5):1060–72.
- 99. Xie J, Yang Y, Gao Y, He J. Cuproptosis: mechanisms and links with cancers. Mol Cancer. 2023;22(1):46.
- Gong Y, Fan Z, Luo G, Yang C, Huang Q, Fan K, et al. The role of necroptosis in cancer biology and therapy. Mol Cancer. 2019;18(1):100.
- 101. Wei X, Xie F, Zhou X, Wu Y, Yan H, Liu T, et al. Role of pyroptosis in inflammation and cancer. Cell Mol Immunol. 2022;19(9):971–92.
- 102. Shchors K, Massaras A, Hanahan D. Dual targeting of the autophagic regulatory circuitry in gliomas with repurposed drugs elicits cell-lethal autophagy and therapeutic benefit. Cancer Cell. 2015;28(4):456–71.
- 103. Bhutia SK, Mukhopadhyay S, Sinha N, Das DN, Panda PK, Patra SK, et al. Autophagy: cancer's friend or foe? Adv Cancer Res. 2013;118:61–95.
- Smithuis F, Kyaw MK, Phe O, Win T, Aung PP, Oo APP, et al. Effectiveness of five artemisinin combination regimens with or without primaquine in uncomplicated falciparum malaria: an open-label randomised trial. Lancet Infect Dis. 2010;10(10):673–81.
- 105. Wong YK, Xu C, Kalesh KA, He Y, Lin Q, Wong WF, et al. Artemisinin as an anticancer drug: recent advances in target profiling and mechanisms of action. Med Res Rev. 2017;37(6):1492–517.
- Yang N-D, Tan S-H, Ng S, Shi Y, Zhou J, Tan KSW, et al. Artesunate induces cell death in human cancer cells via enhancing lysosomal function and lysosomal degradation of ferritin. J Biol Chem. 2014;289(48):33425–41.
- 107. Chen G-Q, Benthani FA, Wu J, Liang D, Bian Z-X, Jiang X. Artemisinin compounds sensitize cancer cells to ferroptosis by regulating iron homeostasis. Cell Death Differ. 2020;27(1):242–54.
- 108. Du J, Wang T, Li Y, Zhou Y, Wang X, Yu X, et al. DHA inhibits proliferation and induces ferroptosis of leukemia cells through autophagy dependent degradation of ferritin. Free Radical Biol Med. 2019;131:356–69.
- Zhang J, Guo L, Zhou X, Dong F, Li L, Cheng Z, et al. Dihydroartemisinin induces endothelial cell anoikis through the activation of the JNK signaling pathway Corrigendum. Oncol Lett. 2016;12(3):1896–900.
- 110. Janku F, McConkey DJ, Hong DS, Kurzrock R. Autophagy as a target for anticancer therapy. Nat Rev Clin Oncol. 2011;8(9):528–39.
- 111. Wolpin BM, Rubinson DA, Wang X, Chan JA, Cleary JM, Enzinger PC, et al. Phase II and pharmacodynamic study of autophagy inhibition using hydroxychloroquine in patients with metastatic pancreatic adenocarcinoma. Oncologist. 2014;19(6):637–8.
- 112. Pantziarka P, Vandeborne L, Bouche G. A database of drug repurposing clinical trials in oncology. Front Pharmacol. 2021. https://doi.org/10.3389/fphar.2021.790952.
- 113. Brugmans JP, Thienpont DC, van Wijngaarden I, Vanparijs OF, Schuermans VL, Lauwers HL. Mebendazole in enterobiasis. Radiochemical and pilot clinical study in 1,278 subjects. Jama. 1971;217(3):313–6.

- 114. Sasaki J, Ramesh R, Chada S, Gomyo Y, Roth JA, Mukhopadhyay T. The anthelmintic drug mebendazole induces mitotic arrest and apoptosis by depolymerizing tubulin in non-small cell lung cancer cells. Mol Cancer Ther. 2002;1(13):1201–9.
- 115. Guerini AE, Triggiani L, Maddalo M, Bonù ML, Frassine F, Baiguini A, et al. Mebendazole as a candidate for drug repurposing in oncology: an extensive review of current literature. Cancers. 2019. https://doi.org/10.3390/cancers11091284.
- Kralova V, Hanušová V, Caltová K, Špaček P, Hochmalová M, Skálová L, Rudolf E. Flubendazole and mebendazole impair migration and epithelial to mesenchymal transition in oral cell lines. Chem Biol Interact. 2018;293:124–32.
- 117. Joe NS, Godet I, Milki N, Ain NUI, Oza HH, Riggins GJ, Gilkes DM. Mebendazole prevents distant organ metastases in part by decreasing ITGβ4 expression and cancer stemness. Breast Cancer Res. 2022;24(1):98.
- Zhang J, Wei W, Zhong Q, Feng K, Yang R, Jiang Q. Budding uninhibited by benzimidazoles 1 promotes cell proliferation, invasion, and epithelial-mesenchymal transition via the Wnt/β-catenin signaling in glioblastoma. Heliyon. 2023;9(6):e16996.
- 119. Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. Cell Metab. 2016;23(1):27-47.
- 120. Sainero-Alcolado L, Liaño-Pons J, Ruiz-Pérez MV, Arsenian-Henriksson M. Targeting mitochondrial metabolism for precision medicine in cancer. Cell Death Differ. 2022;29(7):1304–17.
- 121. Pollak M. Overcoming drug development bottlenecks with repurposing: repurposing biguanides to target energy metabolism for cancer treatment. Nat Med. 2014;20(6):591–3.
- 122. Rao Kondapally Seshasai S, Kaptoge S, Thompson A, Di Angelantonio E, Gao P, Sarwar N, et al. Diabetes mellitus, fasting glucose, and risk of cause-specific death. N Engl J Med. 2011;364(9):829–41.
- 123. Gallagher EJ, LeRoith D. Obesity and diabetes: the increased risk of cancer and cancer-related mortality. Physiol Rev. 2015;95(3):727–48.
- 124. Pernicova I, Korbonits M. Metformin—mode of action and clinical implications for diabetes and cancer. Nat Rev Endocrinol. 2014;10(3):143–56.
- 125. Lord SR, Harris AL. Is it still worth pursuing the repurposing of metformin as a cancer therapeutic? Br J Cancer. 2023;128(6):958–66.
- 126. De A, Kuppusamy G. Metformin in breast cancer: preclinical and clinical evidence. Curr Probl Cancer. 2020;44(1):100488.
- 127. Fendt S-M, Bell EL, Keibler MA, Davidson SM, Wirth GJ, Fiske B, et al. Metformin decreases glucose oxidation and increases the dependency of prostate cancer cells on reductive glutamine metabolism. Can Res. 2013;73(14):4429–38.
- 128. Buzzai M, Jones RG, Amaravadi RK, Lum JJ, DeBerardinis RJ, Zhao F, et al. Systemic treatment with the antidiabetic drug metformin selectively impairs p53-deficient tumor cell growth. Can Res. 2007;67(14):6745–52.
- 129. Blandino G, Valerio M, Cioce M, Mori F, Casadei L, Pulito C, et al. Metformin elicits anticancer effects through the sequential modulation of DICER and c-MYC. Nat Commun. 2012;3(1):865.
- 130. Hald J, Jacobsen E. A drug sensitising the organism to ethyl alcohol. Lancet. 1948;252(6539):1001-4.
- 131. Veverka KA, Johnson KL, Mays DC, Lipsky JJ, Naylor S. Inhibition of aldehyde dehydrogenase by disulfiram and its metabolite methyl diethylthiocarbamoyl-sulfoxide. Biochem Pharmacol. 1997;53(4):511–8.
- 132. Dorokhov YL, Sheshukova EV, Bialik TE, Komarova TV. Human endogenous formaldehyde as an anticancer metabolite: its oxidation downregulation may be a means of improving therapy. BioEssays. 2018;40(12):1800136.
- 133. Falls-Hubert KC, Butler AL, Gui K, Anderson M, Li M, Stolwijk JM, et al. Disulfiram causes selective hypoxic cancer cell toxicity and radio-chemo-sensitization via redox cycling of copper. Free Radical Biol Med. 2020;150:1–11.
- 134. Skrott Z, Mistrik M, Andersen KK, Friis S, Majera D, Gursky J, et al. Alcohol-abuse drug disulfiram targets cancer via p97 segregase adaptor NPL4. Nature. 2017;552(7684):194–9.
- 135. Mohme M, Riethdorf S, Pantel K. Circulating and disseminated tumour cells—mechanisms of immune surveillance and escape. Nat Rev Clin Oncol. 2017;14(3):155–67.
- 136. Teng MW, Swann JB, Koebel CM, Schreiber RD, Smyth MJ. Immune-mediated dormancy: an equilibrium with cancer. J Leucocyte Biol. 2008;84(4):988–93.
- 137. Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from immunosurveillance to tumor escape. Nat Immunol. 2002;3(11):991–8.
- Chen DS, Mellman I. Elements of cancer immunity and the cancer-immune set point. Nature. 2017;541(7637):321–30.
- 139. Mulder WJM, Ochando J, Joosten LAB, Fayad ZA, Netea MG. Therapeutic targeting of trained immunity. Nat Rev Drug Discov. 2019;18(7):553–66.
- 140. Melero I, Gato M, Shekarian T, Aznar A, Valsesia-Wittmann S, Caux C, et al. Repurposing infectious disease vaccines for intratumoral immunotherapy. J Immunother Cancer. 2020. https://doi.org/10.1136/jitc-2019-000443.
- 141. Shekarian T, Sivado E, Jallas AC, Depil S, Kielbassa J, Janoueix-Lerosey I, et al. Repurposing rotavirus vaccines for intratumoral immunotherapy can overcome resistance to immune checkpoint blockade. Sci Transl Med. 2019. https://doi.org/10.1126/scitranslmed.aat5025.
- 142. Aznar MA, Molina C, Teijeira A, Rodriguez I, Azpilikueta A, Garasa S, et al. Repurposing the yellow fever vaccine for intratumoral immunotherapy. EMBO Mol Med. 2020;12(1):e10375.
- 143. Mastrangelo G, Rossi CR, Pfahlberg A, Marzia V, Barba A, Baldo M, et al. Is there a relationship between influenza vaccinations and risk of melanoma? A population-based case-control study. Eur J Epidemiol. 2000;16(9):777–82.
- 144. Sherr CJ, McCormick F. The RB and p53 pathways in cancer. Cancer Cell. 2002;2(2):103–12.
- 145. Van Dyke K, Lantz C, Szustkiewicz C. Quinacrine: mechanisms of antimalarial action. Science.
- 1970;169(3944):492–3.
 Gurova KV, Hill JE, Guo C, Prokvolit A, Burdelya LG, Samoylova E, et al. Small molecules that reactivate p53 in renal cell carcinoma reveal a NF-kappaB-dependent mechanism of p53 suppression in tumors. Proc Natl Acad Sci U S A.
- 2005;102(48):17448–53. 147. Batchu RB, Gruzdyn OV, Bryant CS, Qazi AM, Kumar S, Chamala S, et al. Ritonavir-mediated induction of apoptosis
- (47. Batchu RB, Gruzdyn OV, Bryant CS, Qazi AM, Kumar S, Chamala S, et al. Ritonavir-mediated induction of apoptosis in pancreatic cancer occurs via the RB/E2F-1 and AKT pathways. Pharmaceuticals. 2014;7(1):46–57.

- 148. Mandal CC, Ghosh-Choudhury N, Yoneda T, Choudhury GG, Ghosh-Choudhury N. Simvastatin prevents skeletal metastasis of breast cancer by an antagonistic interplay between p53 and CD44. J Biol Chem. 2011;286(13):11314–27.
- 149. Miyajima C, Hayakawa Y, Inoue Y, Nagasaka M, Hayashi H. HMG-CoA reductase inhibitor statins activate the transcriptional activity of p53 by regulating the expression of TAZ. Pharmaceuticals. 2022;15(8):1015.
- 150. Bergers G, Benjamin LE. Tumorigenesis and the angiogenic switch. Nat Rev Cancer. 2003;3(6):401–10.
- 151. Albini A, Tosetti F, Li VW, Noonan DM, Li WW. Cancer prevention by targeting angiogenesis. Nat Rev Clin Oncol. 2012;9(9):498–509.
- 152. Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. Nat Rev Cancer. 2008;8(8):592–603.
- 153. Schein CH. Repurposing approved drugs on the pathway to novel therapies. Med Res Rev. 2020;40(2):586–605.
- Eleutherakis-Papaiakovou V, Bamias A, Dimopoulos MA. Thalidomide in cancer medicine. Ann Oncol. 2004;15(8):1151–60.
- Yabu T, Tomimoto H, Taguchi Y, Yamaoka S, Igarashi Y, Okazaki T. Thalidomide-induced antiangiogenic action is mediated by ceramide through depletion of VEGF receptors, and is antagonized by sphingosine-1-phosphate. Blood. 2005;106(1):125–34.
- 156. Richardson P, Anderson K. Thalidomide and dexamethasone: a new standard of care for initial therapy in multiple myeloma. J Clin Oncol. 2006;24(3):334–6.
- 157. Como JA, Dismukes WE. Oral azole drugs as systemic antifungal therapy. N Engl J Med. 1994;330(4):263–72.
- 158. Tsubamoto H, Ueda T, Inoue K, Sakata K, Shibahara H, Sonoda T. Repurposing itraconazole as an anticancer agent. Oncol Lett. 2017;14(2):1240–6.
- 159. Wang SJ, Sun B, Cheng ZX, Zhou HX, Gao Y, Kong R, et al. Dihydroartemisinin inhibits angiogenesis in pancreatic cancer by targeting the NF-κB pathway. Cancer Chemother Pharmacol. 2011;68(6):1421–30.
- 160. Bhise NS, Shmueli RB, Sunshine JC, Tzeng SY, Green JJ. Drug delivery strategies for therapeutic angiogenesis and antiangiogenesis. Expert Opin Drug Deliv. 2011;8(4):485–504.
- 161. Świtalska M, Filip-Psurska B, Milczarek M, Psurski M, Moszyńska A, Dąbrowska AM, et al. Combined anticancer therapy with imidazoacridinone analogue C-1305 and paclitaxel in human lung and colon cancer xenografts modulation of tumour angiogenesis. J Cell Mol Med. 2022;26(14):3950–64.
- Diakos CI, Charles KA, McMillan DC, Clarke SJ. Cancer-related inflammation and treatment effectiveness. Lancet Oncol. 2014;15(11):e493-503.
- Greten FR, Grivennikov SI. Inflammation and cancer: triggers, mechanisms, and consequences. Immunity. 2019;51(1):27–41.
- 164. Wang J, Wu K, Bai F, Zhai H, Xie H, Du Y, et al. Celecoxib could reverse the hypoxia-induced Angiopoietin-2 upregulation in gastric cancer. Cancer Lett. 2006;242(1):20–7.
- 165. Lin XM, Li S, Zhou C, Li RZ, Wang H, Luo W, et al. Cisplatin induces chemoresistance through the PTGS2-mediated anti-apoptosis in gastric cancer. Int J Biochem Cell Biol. 2019;116:105610.
- Vergani E, Dugo M, Cossa M, Frigerio S, Di Guardo L, Gallino G, et al. miR-146a-5p impairs melanoma resistance to kinase inhibitors by targeting COX2 and regulating NFkB-mediated inflammatory mediators. Cell Commun Signal. 2020;18(1):156.
- 167. Guo Q, Li Q, Wang J, Liu M, Wang Y, Chen Z, et al. A comprehensive evaluation of clinical efficacy and safety of celecoxib in combination with chemotherapy in metastatic or postoperative recurrent gastric cancer patients: A preliminary, three-center, clinical trial study. Medicine. 2019;98(27):e16234.
- 168. Gasic GJ, Gasic TB, Galanti N, Johnson T, Murphy S. Platelet-tumor-cell interactions in mice. The role of platelets in the spread of malignant disease. Int J Cancer. 1973;11(3):704–18.
- 169. Li M, Lotan R, Levin B, Tahara E, Lippman SM, Xu X-C. Aspirin induction of apoptosis in esophageal cancer: a potential for chemoprevention 1. Cancer Epidemiol Biomark Prev. 2000;9(6):545–9.
- Kumar D, Rahman H, Tyagi E, Liu T, Li C, Lu R, et al. Aspirin suppresses PGE2 and activates AMP kinase to inhibit melanoma cell motility, pigmentation, and selective tumor growth in vivo. Cancer Prev Res. 2018;11(10):629–42.
- 171. Barnard ME, Beeghly-Fadiel A, Milne GL, Akam EY, Chan AT, Eliassen AH, et al. Urinary PGE-M levels and risk of ovarian cancer. Cancer Epidemiol Biomark Prev. 2019;28(11):1845–52.
- 172. Stegeman I, Bossuyt PM, Yu T, Boyd C, Puhan MA. Aspirin for primary prevention of cardiovascular disease and cancer. A benefit and harm analysis. PLoS One. 2015;10(7):e0127194.
- Rothwell PM, Fowkes FG, Belch JF, Ogawa H, Warlow CP, Meade TW. Effect of daily aspirin on long-term risk of death due to cancer: analysis of individual patient data from randomised trials. Lancet. 2011;377(9759):31–41.
- 174. Li P, Wu H, Zhang H, Shi Y, Xu J, Ye Y, et al. Aspirin use after diagnosis but not prediagnosis improves established colorectal cancer survival: a meta-analysis. Gut. 2015;64(9):1419–25.
- Langley RE, Burdett S, Tierney JF, Cafferty F, Parmar MK, Venning G. Aspirin and cancer: has aspirin been overlooked as an adjuvant therapy? Br J Cancer. 2011;105(8):1107–13.
- Xu XR, Yousef GM, Ni H. Cancer and platelet crosstalk: opportunities and challenges for aspirin and other antiplatelet agents. Blood. 2018;131(16):1777–89.
- 177. Zhang X, Feng Y, Liu X, Ma J, Li Y, Wang T, Li X. Beyond a chemopreventive reagent, aspirin is a master regulator of the hallmarks of cancer. J Cancer Res Clin Oncol. 2019;145(6):1387–403.
- 178. Giovannucci E, Rimm EB, Stampfer MJ, Colditz GA, Ascherio A, Willett WC. Aspirin use and the risk for colorectal cancer and adenoma in male health professionals. Ann Internal Med. 1994;121(4):241–6.
- 179. Holmes MD, Chen WY, Li L, Hertzmark E, Spiegelman D, Hankinson SE. Aspirin intake and survival after breast cancer. J Clin Oncol. 2010;28(9):1467–72.
- Jonsson F, Yin L, Lundholm C, Smedby KE, Czene K, Pawitan Y. Low-dose aspirin use and cancer characteristics: a population-based cohort study. Br J Cancer. 2013;109(7):1921–5.
- 181. Skriver C, Maltesen T, Dehlendorff C, Skovlund CW, Schmidt M, Sørensen HT, Friis S. Long-term aspirin use and cancer risk: a 20-year cohort study. J Natl Cancer Inst. 2024;116(4):530–8.
- 182. McNeil JJ, Gibbs P, Orchard SG, Lockery JE, Bernstein WB, Cao Y, et al. Effect of aspirin on cancer incidence and mortality in older adults. J Natl Cancer Inst. 2021;113(3):258–65.

- Regulska K, Regulski M, Karolak B, Michalak M, Murias M, Stanisz B. Beyond the boundaries of cardiology: still untapped anticancer properties of the cardiovascular system-related drugs. Pharmacol Res. 2019;147:104326.
- 184. Shah P, Garris R, Abboud R, Vasudev R, Patel H, Doshi R, et al. Meta-analysis comparing usefulness of beta blockers to preserve left ventricular function during anthracycline therapy. Am J Cardiol. 2019;124(5):789–94.
- 185. Spini A, Roberto G, Gini R, Bartolini C, Bazzani L, Donnini S, et al. Evidence of β-blockers drug repurposing for the treatment of triple negative breast cancer: a systematic review. Neoplasma. 2019;66(6):963–70.
- 186. Hagen R, Ghareeb E, Jalali O, Zinn Z. Infantile hemangiomas: what have we learned from propranolol? Curr Opin Pediatr. 2018;30(4):499–504.
- Wagner MJ, Cranmer LD, Loggers ET, Pollack SM. Propranolol for the treatment of vascular sarcomas. J Exp Pharmacol. 2018;10:51–8.
- 188. Breckenridge A, Jacob R. Overcoming the legal and regulatory barriers to drug repurposing. Nat Rev Drug Discovery. 2019;18(1):1–2.
- 189. Pantziarka P, Bouche G, Meheus L, Sukhatme V, Sukhatme V, Vikas P. The repurposing drugs in oncology (ReDO) project. Ecancermedicalscience. 2014;8(442):2014.
- 190. Cha Y, Erez T, Reynolds I, Kumar D, Ross J, Koytiger G, et al. Drug repurposing from the perspective of pharmaceutical companies. Br J Pharmacol. 2018;175(2):168–80.
- 191. Heled Y. Patents v. Statutory exclusivities in biological pharmaceuticals-do we really need both. Mich Telecomm Tech L Rev. 2011;18:419.
- 192. Halabi S. The drug repurposing ecosystem: intellectual property incentives, market exclusivity, and the future of "new" medicines. Yale J Law Technol. 2018;20:1.
- Smith RB. Repositioned drugs: integrating intellectual property and regulatory strategies. Drug Discov Today: Ther Strat. 2011;8(3–4):131–7.
- Radley DC, Finkelstein SN, Stafford RS. Off-label prescribing among office-based physicians. Arch Intern Med. 2006;166(9):1021–6.
- 195. Wieder R, Adam N. Drug repositioning for cancer in the era of AI, big omics, and real-world data. Crit Rev Oncol Hematol. 2022;175:103730.
- 196. Verbaanderd C, Meheus L, Huys I, Pantziarka P. Repurposing drugs in oncology: next steps. Trends in cancer. 2017;3(8):543–6.
- Stewart AK, Jacobus S, Fonseca R, Weiss M, Callander NS, Chanan-Khan AA, Rajkumar SV. Melphalan, prednisone, and thalidomide vs melphalan, prednisone, and lenalidomide (ECOG E1A06) in untreated multiple myeloma. Blood. 2015;126(11):1294–301.
- Zweegman S, van der Holt B, Mellqvist UH, Salomo M, Bos GM, Levin MD, et al. Melphalan, prednisone, and lenalidomide versus melphalan, prednisone, and thalidomide in untreated multiple myeloma. Blood. 2016;127(9):1109–16.
- 199. Tran AA, Prasad V. Drug repurposing for cancer treatments: a well-intentioned, but misguided strategy. Lancet Oncol. 2020;21(9):1134–6.
- Tyrer F, Bhaskaran K, Rutherford MJ. Immortal time bias for life-long conditions in retrospective observational studies using electronic health records. BMC Med Res Methodol. 2022;22(1):86.
- Suissa S, Azoulay L. Metformin and the risk of cancer: time-related biases in observational studies. Diabetes Care. 2012;35(12):2665–73.
- 202. Hammer GP, du Prel J-B, Blettner M. Avoiding bias in observational studies: part 8 in a series of articles on evaluation of scientific publications. Dtsch Arztebl Int. 2009;106(41):664.
- 203. Devita VT Jr, Young RC, Canellos GP. Combination versus single agent chemotherapy: a review of the basis for selection of drug treatment of cancer. 1975;35(1):98–110.
- 204. Day D, Siu LL. Approaches to modernize the combination drug development paradigm. Genome Med. 2016;8:1–14.
- Michel MC, Staskin D. Study designs for evaluation of combination treatment: focus on individual patient benefit. Biomedicines. 2022. https://doi.org/10.3390/biomedicines10020270.
- Lee SJC, Murphy CC, Geiger AM, Gerber DE, Cox JV, Nair R, Skinner CS. Conceptual model for accrual to cancer clinical trials. J Clin Oncol. 2019;37(23):1993.
- Gonzalez-Cao M, Boada A, Teixidó C, Fernandez-Figueras MT, Mayo C, Tresserra F, et al. Fatal gastrointestinal toxicity with ipilimumab after BRAF/MEK inhibitor combination in a melanoma patient achieving pathological complete response. Oncotarget. 2016;7(35):56619.
- Chapman PB, Einhorn LH, Meyers ML, Saxman S, Destro AN, Panageas KS, et al. Phase III multicenter randomized trial of the Dartmouth regimen versus dacarbazine in patients with metastatic melanoma. J Clin Oncol. 1999;17(9):2745.
- 209. Schein CH. Repurposing approved drugs on the pathway to novel therapies. Med Res Rev. 2020;40(2):586–605.
- 210. Bartoszewski R, Sikorski AF. Editorial focus: understanding off-target effects as the key to successful RNAi therapy. Cell Mol Biol Lett. 2019;24(1):69.
- 211. Rationalizing combination therapies. Nature Medicine. 2017;23(10):1113.
- Maziarz M, Stencel A. The failure of drug repurposing for COVID-19 as an effect of excessive hypothesis testing and weak mechanistic evidence. Hist Philos Life Sci. 2022;44(4):47.
- Nguyen N, Jennen D, Kleinjans J. Omics technologies to understand drug toxicity mechanisms. Drug Discov Today. 2022;27(11):103348.
- Verbist B, Klambauer G, Vervoort L, Talloen W, Shkedy Z, Thas O, et al. Using transcriptomics to guide lead optimization in drug discovery projects: lessons learned from the QSTAR project. Drug Discov Today. 2015;20(5):505–13.
- 215. Bartoszewska S, Króliczewski J, Crossman DK, Pogorzelska A, Bagiński M, Collawn JF, Bartoszewski R. Triazoloacridone C-1305 impairs XBP1 splicing by acting as a potential IRE1α endoribonuclease inhibitor. Cell Mol Biol Lett. 2021;26(1):11.

- 216. Króliczewski J, Bartoszewska S, Dudkowska M, Janiszewska D, Biernatowska A, Crossman DK, et al. Utilizing genome-wide mRNA profiling to identify the cytotoxic chemotherapeutic mechanism of triazoloacridone C-1305 as direct microtubule stabilization. Cancers. 2020;12(4):864.
- 217. Pruteanu L-L, Bender A. Using transcriptomics and cell morphology data in drug discovery: the long road to practice. ACS Med Chem Lett. 2023;14(4):386–95.
- 218. Meissner F, Geddes-McAlister J, Mann M, Bantscheff M. The emerging role of mass spectrometry-based proteomics in drug discovery. Nat Rev Drug Discov. 2022;21(9):637–54.
- 219. Page MJ, Amess B, Rohlff C, Stubberfield C, Parekh R. Proteomics: a major new technology for the drug discovery process. Drug Discov Today. 1999;4(2):55–62.
- 220. Frantzi M, Latosinska A, Mischak H. Proteomics in drug development: the dawn of a new era? Proteomics Clin Appl. 2019;13(2):e1800087.
- 221. Pang H, Hu Z. Metabolomics in drug research and development: the recent advances in technologies and applications. Acta Pharm Sin B. 2023;13(8):3238–51.
- 222. Wishart DS. Emerging applications of metabolomics in drug discovery and precision medicine. Nat Rev Drug Discov. 2016;15(7):473–84.
- 223. Van de Sande B, Lee JS, Mutasa-Gottgens E, Naughton B, Bacon W, Manning J, et al. Applications of single-cell RNA sequencing in drug discovery and development. Nat Rev Drug Discov. 2023;22(6):496–520.
- 224. Huang Y, Dong D, Zhang W, Wang R, Lin Y-C-D, Zuo H, et al. DrugRepoBank: a comprehensive database and discovery platform for accelerating drug repositioning. Database. 2024;2024:baae051.
- 225. Masoudi-Sobhanzadeh Y, Omidi Y, Amanlou M, Masoudi-Nejad A. Drug databases and their contributions to drug repurposing. Genomics. 2020;112(2):1087–95.
- Corsello SM, Nagari RT, Spangler RD, Rossen J, Kocak M, Bryan JG, et al. Discovering the anticancer potential of non-oncology drugs by systematic viability profiling. Nat Cancer. 2020;1(2):235–48.
- Ianevski A, Kushnir A, Nader K, Miihkinen M, Xhaard H, Aittokallio T, Tanoli Z. RepurposeDrugs: an interactive webportal and predictive platform for repurposing mono- and combination therapies. Brief Bioinform. 2024. https:// doi.org/10.1093/bib/bbae328.
- 228. Kang BH, Plescia J, Song HY, Meli M, Colombo G, Beebe K, et al. Combinatorial drug design targeting multiple cancer signaling networks controlled by mitochondrial Hsp90. J Clin Invest. 2009;119(3):454–64.
- 229. Czech T, Lalani R, Oyewumi MO. Delivery systems as vital tools in drug repurposing. AAPS PharmSciTech. 2019;20(3):116.
- Séguy L, Groo A-C, Malzert-Fréon A. How nano-engineered delivery systems can help marketed and repurposed drugs in Alzheimer's disease treatment? Drug Discov Today. 2022;27(6):1575–89.
- 231. Weir SJ, DeGennaro LJ, Austin CP. Repurposing approved and abandoned drugs for the treatment and prevention of cancer through public–private partnership. Can Res. 2012;72(5):1055–8.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.